

Customer Name Maeda-Kougyou Japan

218-2 Norimatsu Yahatanishi ward

Kitakyushu City

Japan

Contact

Dr. Khaled Hussein

Customer PO no.

N/A

Test Requested

BS ISO 27447: 2009 Fine ceramics

(advanced ceramics, advanced

technical ceramics) — Test method

for antibacterial activity of

semiconducting photocatalytic

materials (Test method modified due

to client's requests)

Sample Description Textile: non-coated and coated with

Miracle Titanium (Primary and MVX)

Date of Receipt

27th April 2012

Project Number

ASCR0092009

Report Date

5th July 2012



Contents

Description of Test Items		
Introduction	3	
Procedure	3	
Results	ε	
Discussion and Conclusion	8	



Description of Test Items

The following items were tested. The samples to be tested all measured 50 x 50 mm.

Test Item	Product Code	
Treated Textile	ASC002166	
Untreated Textile	ASC002167	

Introduction

The purpose of the project was to ascertain the effect of the MVX coating on *Pseudomonas* aeruginosa viability when applied to a textile following exposure to UV and incandescent light as per the client's request. The test was agreed to be performed in accordance with BS ISO 27447: 2009 Fine ceramics (advanced ceramics, advanced technical ceramics) — Test method for antibacterial activity of semiconducting photocatalytic materials. As the client's requests included alterations in the bacterial to be used, some alterations were made, however these have been subsequently noted.

Procedure

The experimental procedure was performed in accordance with BS ISO 27447: 2009 Fine ceramics (advanced ceramics, advanced technical ceramics) — Test method for antibacterial activity of semiconducting photocatalytic materials, with alterations made to accommodate the client's sample and microbiological challenge specifications. The method for analysis was as follows:

Textile Samples (Glass adhesion method)

- Prior to inoculation all samples and coverslips were sterilised by autoclaving at 121°C for 15 minutes.
- 2. P. aeruginosa was grown in Tryptic Soy Broth under aerobic conditions at 37°C for 18 hours.
- 3. The concentration of bacteria was adjusted to a target concentration of 1 x 10⁵ cells ml⁻¹ using 1/500 nutrient broth.
- 4. Specimens were individually placed in sterile petri dishes. In order to preserve moisture a sterile filter paper was moistened with sterile water with the specimens to be tested separated by a glass slide.



- 5. A 150 μl aliquot of the adjusted bacterial suspension was placed on the surface of the textile samples and a glass slide was placed on top to press the bacterial suspension uniformly under the glass.
- Treated and untreated samples were kept in a dark place or exposed to the light source as specified in the standard with the additional incandescent light source as per the client's request.
- 7. Three untreated samples were immediately washed in 20 ml PBS. A 2 ml aliquot of this washout was serially diluted in sterile phosphate buffered saline (PBS).
- 8. A 500 μl aliquot of the neat washout and the serially diluted samples was plated in duplicate on sterile petri-dishes.
- 9. Approximately 15 20 mls of Tryptic Soy Agar was placed into each petri dish in order to enumerate viable colony forming units by the pour plate method following incubation at 37°C for 24 48 hours. Where the number of cfu exceeded 300 the plates were recorded as TNTC (Too numerous to count).

Satisfaction of criteria for a valid test and calculations

The test requirement fulfilment validation follows the raw data in the results section (see below). In addition the results expressing photocatalyst antibacterial activity value on textiles (S_L) and the photocatalyst antibacterial activity value with UV and incandescent light irradiation on textiles (ΔS).

Glass adhesion method

 $M = P \times 20$

M is the number of cells of viable bacteria

P is the bacteria concentration (cells/ml)

20 is the quantity of PBS used for washout (ml)

Propagation values for validation of conditions for a valid test

 $F_{BL} = M_{BL} - M_{BA}$

 F_{BL} is the growth value after light exposure

 $M_{\rm BL}$ is the average logarithmic value of the number of bacteria for 3 non treated specimens after light exposure

 $M_{\rm BA}$ is the average logarithmic value of the number of viable bacteria for three non treated specimens just after inoculation



 $F_{BD} = M_{BD} - M_{BA}$

 F_{BD} is the growth value after being kept in a dark place

 $M_{\rm BD}$ is the average logarithmic value of the number of viable bacteria for three non treated specimens after being kept in a dark place

Photocatalyst antibacterial activity value calculation

 $S_L = M_{BL} - M_L$

S_L is the photocatalyst antibacterial activity value after light exposure

 M_L is the average logarithmic value of the number of viable bacteria for 3 photocatalytic treated specimens after light exposure

 $\Delta S = (M_{BL} - M_L) - (M_{BD} - M_D)$

ΔS is the photocatalyst antibacterial value with light exposure

M_D is the average logarithmic value of the number of viable bacteria for three photocatalytic treated specimens after being kept in a dark place



Results

Pseudomonas aeuruginosa Textile

Sample description	Dilution	Colony count	Number of viable bacteria recovered per specimen	Log values
Untreated 1	1 x 10°	TNTC*	7600	3.88081359
t = 0	1 x 10 ⁻¹	17, 21		3.00061333
Untreated 2	1 x 10 ⁰	TNTC	7000	3.84509804
t=0	1 x 10 ⁻¹	16, 19		3.64303604
Untreated 3	1 x 10°	TNTC	8200	3.91381385
t=0	1 x 10 ⁻¹	21, 20	8200	3.91301303
Light	1 x 10°	TNTC	9400	3.97312785
untreated 1	1 x 10 ⁻¹	19, 28		3.97312763
Light	1 x 10°	TNTC	- 7800	3.8920946
untreated 2	1 x 10 ⁻¹	20, 19		3.6920940
Light	1 x 10°	TNTC	7400	3.86923172
untreated 3	1 x 10 ⁻¹	19, 18		3.80923172
Light	1 x 10°	1, 0	40	1.60205999
treated 1	1 x 10 ⁻¹	0, 0		1.00203333
Light	1 x 10°	0, 0	0	0
treated 2	1 x 10 ⁻¹	0, 0		- 0
Light	1 x 10°	0, 0	0	0
treated 3	1 x 10 ⁻¹	0, 0		O
Dark	1 x 10 ⁰	TNTC	4600	3.66275783
untreated 1	1 x 10 ⁻¹	10, 13		3,002/3/63
Dark	1 x 10°	TNTC	8200	3.91381385
untreated 2	1 x 10 ⁻¹ 17, 24	3.31301303		
Dark	1 x 10°	TNTC	13200	4.12057393
untreated 3	1 x 10 ⁻¹	16, 17		7.12037333



Dark treated	1 x 10°	TNTC	3200	3.50514998
	1 x 10 ⁻¹	8, 8		
Dark treated	1 x 10°	x 10 ⁰ TNTC 4000	3.60205999	
2	1 x 10 ⁻¹	8, 12	4000	3.00203333
Dark treated	1 x 10°	TNTC	1000	3
3	1 x 10 ⁻¹	2, 3	1000	3

*TNTC: Too Numerous To Count

Test requirement fulfilment validation

1. F_{BL} = 3.911 – 3.873 = 0.038 F_{BL} is greater than 0 therefore parameter is validated

2. F_{BD} = 3.898 – 3.873 = 0.025 F_{BD} is greater than 0 therefore parameter is validated

$$S_L$$
 = 3.911 - 0.533 = 3.378
 ΔS = (3.911 - 0.533) - (3.898 - 3.369)
= 3.378 - 0.529
= 2.849



Discussion and Conclusion

In accordance with the wishes of the client the ISO 22447:2009 procedure was modified slightly as the surface to be tested consisted of textile samples. In addition, as agreed with the client Pseudomonas aeruginosa was used for analysing the effect of the coatings against viability of the bacteria of interest when exposed to light.

In understanding the data it must be noted that S_L values account for the reduction of viability caused by the exposure of the treated surfaces to light on textiles. In contrast, ΔS values address the reduction of bacterial viability caused by the coating becoming light activated while accounting for the reduction in viability caused by the same coatings in a dark environment. From the data presented here it is clear that in all cases the coatings resulted in a reduction in bacterial viability even when stored in a dark place. It is therefore clear that the coatings in the absence of light have a bactericidal effect.

As per the client's request P. aeruginosa NCTC10622 was tested on the textile surfaces using the glass adhesion method. The data demonstrated that the coatings were highly effective against P. aeruginosa demonstrating a ΔS value of 2.849 with the criteria for a valid test fulfilled.

In conclusion the coatings appear to be very effective at reducing the viability of P. aeruginosa on textile.

Report Compiled by:

John Fallon

John Fallon PhD Senior Scientific Officer Report Reviewed by:

Digitally signed by Maire For Maire Fox

document Date: 2012.07.05 12:05:10 +01'00'

Máire Fox MSc **Laboratory Manger**

*** End of Report***